



Enantiomerically pure *N*-Boc- and *N*-benzoyl-(*S*)-phenylglycinals

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Abstract—Enantiomerically pure *N*-Boc- and *N*-benzoyl-(*S*)-phenylglycinals were prepared by oxidation of the respective alcohols with Dess–Martin periodinane. The glycinals were phosphonylated with lithium *O,O*-dimethyl phosphonate at -70°C or $(\text{MeO})_2\text{POTMS}$ at -20°C without racemisation. In the presence of 10 mol% of NEt_3 at 20°C the aldehydes racemised instantaneously, while it took a few hours for the re racemisation processes to reach completion after addition of 1 mol% of NEt_3 . © 2002 Published by Elsevier Science Ltd.

1. Introduction

N-Benzoyl- and *N*-Boc-phenylglycinals, **1** and **2**, belong to a group of α -amino aldehydes¹ usually considered as chemically unstable and easily racemised compounds. Since they have been employed in syntheses of several important natural products^{2–4} including paclitaxel^{5–10} and paclitaxel analogues,^{11–14} the preparation of enantiomerically pure **1** and **2** is an important goal. However, in almost all synthetic applications of **1** and **2** the materials prepared in situ were immediately used in further transformations. An attempt of purification by vacuum distillation of *N*-Boc-*D*-phenylglycinal was described by Shioiri.⁴

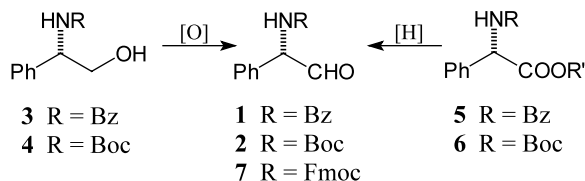
In general, α -amino aldehydes **1** and **2** were synthesised (Scheme 1) by oxidation of the respective α -amino alcohols **3** and **4**^{2–5,10,11,15} and by reduction of esters **5** and **6**^{6,7,9} or Weinreb amides.⁸ Other methods are also

known.^{16,17} Oxidations were performed under Swern conditions using ethyldiisopropylamine^{2,5,10,11} or with sulphur trioxide–pyridine complex.⁴ The crude aldehydes were further subjected to nucleophilic additions, and their e.e.'s were estimated by the analysis of the enantiomeric purities of the addition products. DIBAL-H reduction of esters **5** and **6** elaborated by Dondoni⁹ led to the aldehydes **1** and **2**, and their e.e.'s were established as 84 and 90%, respectively, after NaBH_4 reduction to the known phenylglycinols **3** and **4**.

The recent achievements of Myers in the synthesis of enantiomerically pure Fmoc-phenylglycinal^{18–20} **7** and scarce examples of the application of the Dess–Martin reagent to the oxidation of **4**^{5,15} prompted us to investigate the preparation of enantiomerically pure **1** and **2** and their racemisation-free transformation into the respective α -hydroxy phosphonates.

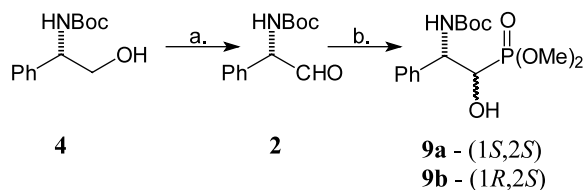
2. Results and discussion

(*S*)-Phenylglycinol **8** was obtained from *L*-phenylglycine according to the described procedure.²¹ In the presence of Boc_2O **8** was transformed into *N*-Boc derivative **4**,²² while benzylation led to *N*-benzoyl-(*S*)-phenylglycinol **3**.²² Oxidation of **4** with the Swern reagent^{23,24} was initially attempted. Despite the poor solubility of **4** in dichloromethane at low temperature, the aldehyde formed in situ was treated with lithium *O,O*-dimethyl phosphonate at -70°C to give a 74:26 mixture of the phosphonates **9a** and **9b** (Scheme 2).¹⁴ The e.e. of **9a** and **9b** was established as only 76% by the ³¹P NMR analysis of the ω -camphanate derivatives.¹⁴



Scheme 1.

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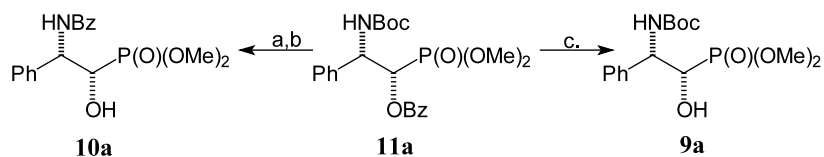
Scheme 2. Reagents and conditions: (a) Swern (NEt₃) or Dess–Martin reagents; (b) LiP(O)(OMe)₂, –70°C or (MeO)₂POTMS, –20°C and Bu₄NF.

However, when Dess–Martin periodinane^{25–28} was applied, the aldehyde **2** was obtained as a white amorphous solid after crystallisation from CH₂Cl₂–ether–hexanes at –15°C. Reduction of a sample of **2** with NaBH₄ at 0°C⁹ led to **4**, which was esterified with (*S*)-*O*-methylmandelic acid and found to be enantiomerically pure by ¹H NMR analysis of the reaction mixture. Phosphonylation of **2** was accomplished with (MeO)₂POTMS at –20°C, and after desilylation with Bu₄NF, a 83:17 mixture of **9a** and **9b** was produced. The 100% e.e. of the phosphonates obtained was proved by the ³¹P NMR analysis of their ω-camphanates.

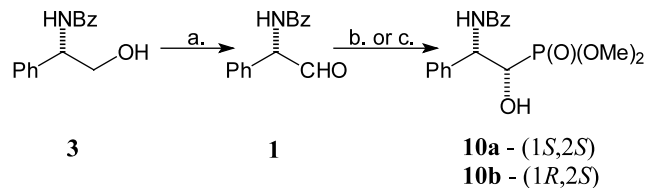
In order to verify whether lithium *O,O*-dimethyl phosphonate could cause racemisation of **2** at –70°C, the aldehyde prepared after Dess–Martin oxidation of **4** was subjected to phosphonylation under these conditions. The 100% e.e. of the addition products was proved by the ³¹P NMR spectroscopy. Enantiomerically pure phosphonate analogues of paclitaxel and docetaxel side chain **10a** and **9a**, respectively, were next obtained as follows. After benzylation of the crude mixture of diastereoisomers **9a** and **9b**, the benzoate **11a** was separated in 31% yield after silica gel chromatography. The ester **11a** was subjected to a two-step one-pot procedure¹⁴ to produce the phosphonate **10a** in 82% yield, while ammonolysis²⁹ of **11a** gave **9a** in 40% yield (Scheme 3). Both phosphonates were shown to be 100% e.e. by HPLC.

Dess–Martin periodinane oxidation of *N*-benzoyl-(*S*)-phenylglycinol **3** afforded enantiomerically pure aldehyde **1** as a white amorphous solid after crystallisation from CH₂Cl₂–ether–hexanes at –15°C. The aldehyde was reacted with either lithium *O,O*-dimethyl phosphonate at –70°C or dimethyl (trimethylsilyl)phosphite at –20°C to give 1:1 and 87:13 mixtures of the phosphonates **10a** and **10b**, respectively (Scheme 4).

Again, 100% e.e.'s of the phosphonates formed were established by the ³¹P NMR spectroscopy after esterifi-



Scheme 3. Reagents and conditions: (a) HCl–AcOEt, CH₂Cl₂; (b) NEt₃; (c) NH₃ aq., MeOH.



Scheme 4. Reagents and conditions: (a) Dess–Martin reagent; (b) LiP(O)(OMe)₂, –70°C; (c) (MeO)₂POTMS, –20°C, then Bu₄NF.

cation with ω-camphanyl chloride¹⁴ and further proved by HPLC.

After successful preparation of the enantiomerically pure *N*-Boc- and *N*-benzoyl-(*S*)-phenylglycinols, **1** and **2**, we addressed the problem of configurational stability of these compounds. Even crude products isolated after oxidation with Dess–Martin periodinane retained their 100% enantiomeric purity when left at 20°C for 24 h. At 0°C the purified aldehydes kept their configurational integrity for at least 3 months. However, at 20°C in the presence of 10 mol% of triethylamine, the purified aldehydes underwent complete racemisation instantaneously. The rates of racemisation of (*S*)-**1** and (*S*)-**2** were qualitatively followed at 20°C after addition of 1 mol% of NEt₃ (Table 1).

Table 1. The rate of racemisation of the aldehydes (*S*)-**1** and (*S*)-**2**

Time (h)	0	1	2	3	24
(<i>S</i>)- 1	90 ^a	38	32	30	0
(<i>S</i>)- 2	100	55	50	40	0

^a E.e. from ¹H NMR spectroscopy.

Undoubtedly, the lower enantiomeric purity of the aldehyde **2** obtained by oxidation of **4** with Swern reagent was caused by the presence of triethylamine.

3. Conclusions

Dess–Martin periodinane was successfully applied to a racemisation-free oxidation of *N*-Boc- and *N*-benzoyl-(*S*)-phenylglycinols, while the Swern reagent caused some racemisation even at –60°C. The conditions of phosphonylation of enantiomerically pure *N*-Boc- and *N*-benzoyl-(*S*)-phenylglycinols were elaborated, namely with lithium *O,O*-dimethyl phosphonate at –70°C or with dimethyl(trimethylsilyl)phosphite at –20°C. In the presence of 1 mol% of NEt₃ the aldehydes racemised

within a few hours at 20°C, while after addition of 10 mol% of NEt₃, racemisation occurred instantaneously.

4. Experimental

¹H, ¹³C and ³¹P NMR spectra were taken in CDCl₃ on the Varian Mercury-300 spectrometer at 300, 75.5 and 121.5 MHz, respectively. IR spectral data were measured on an Infinity MI-60 FT-IR spectrometer. Melting points were determined on a Boetius apparatus and are uncorrected. Elemental analysis were performed by the Microanalytical Laboratory of this Faculty on a Perkin–Elmer PE 2400 CHNS analyser. Polarimetric measurements were conducted on a Perkin–Elmer 241 MC apparatus. HPLC analyses were carried out on a LDC Analytical apparatus (column: Chiracel OD, 0.46 cm ϕ ×25 cm; detection: UV at 256 nm; isopropanol–hexanes, 1:9; 1 mL/min; rt).

The following adsorbents were used: column chromatography, Merck silica gel 60 (70–230 mesh); analytical TLC, Merck TLC plastic sheets silica gel 60 F₂₅₄. TLC plates were developed in various ethyl acetate–hexanes or chloroform–methanol solvent systems. Visualisation of spots was effected with iodine vapours.

4.1. *N*-Benzoyl- and *N*-Boc-(*S*)-phenylglycinals, **1** and **2**

To a solution of amino alcohol **3** or **4** (482 mg or 475 mg, 2.00 mmol) in water-saturated CH₂Cl₂ (7.5 mL) was added Dess–Martin periodinane (1.78 g, 4.20 mmol). The suspension was stirred at 23°C for 30 min while water-saturated CH₂Cl₂ (2×3 mL) was added every 15 min. After disappearance of the starting alcohols (TLC), ethyl ether (6.7 mL) was added followed by a solution of sodium thiosulphate (5.46 g) in 80% saturated sodium bicarbonate (7.3 mL). Organic layer was separated, and the aqueous phase was extracted with ether (15 mL). Organic solution was washed with saturated aqueous NaHCO₃ (10 mL), water (2×10 mL) and brine (2×10 mL). After drying over MgSO₄ at 0°C, the solvents were partially evaporated (water bath below 20°C). The residue was triturated with hexanes until turbidity and left at –15°C for 1 h (in the case of **1**) or overnight (in the case of **2**). Solvents were removed, solid aldehydes were washed with a 4:1 hexane–ether mixture and dried in a stream of argon at rt.

4.1.1. *N*-Benzoyl-(*S*)-phenylglycinal, (*S*)-1**.** Yield 230 mg (48%). Mp: 121–122°C; $[\alpha]_{\text{D}}^{20}$ +269 (*c* 0.8 in CH₂Cl₂); $[\alpha]_{\text{D}}^{20}$ +260 (*c* 0.85 in CHCl₃).

4.1.2. *N*-Boc-(*S*)-phenylglycinal, (*S*)-2**.** Yield 250 mg (53%). Mp: 75.9–76.4°C, lit.⁴ 55–56°C; $[\alpha]_{\text{D}}^{20}$ +272 (*c* 0.9 in CH₂Cl₂), lit.⁴ $[\alpha]_{\text{D}}^{19.5}$ –95.5 (*c* 0.53 in CH₂Cl₂) for (*R*)-**2**; $[\alpha]_{\text{D}}^{20}$ +308 (*c* 0.75 in CHCl₃).

4.2. General procedure for the e.e. determination of (*S*)-**1** and (*S*)-**2**

4.2.1. Method 1: by ¹H NMR spectroscopy. A sample (50 mg, 0.20 mmol) of the aldehyde **1** or **2** was dissolved in methanol (1 mL), cooled to 0°C and NaBH₄ (12 mg, 0.32 mmol) was added. After 30 min the reaction mixture was allowed to reach rt and was neutralised to pH 7 with 1N HCl. Methanol was evaporated, the residue was suspended in CH₂Cl₂ (5 mL) and anhydrous MgSO₄ (1 g) was added. The solids were removed and the solution was evaporated to give crude phenylglycinals **3** and **4** quantitatively.

A sample of **3** or **4** (10 mg, 0.041 mmol) was dissolved in CH₂Cl₂ (1.0 mL) and (*S*)-*O*-methylmandelic acid (8.0 mg, 0.046 mmol) was added followed by DCC (9.0 mg, 0.046 mmol) and a crystal of DMAP. After 2 h at rt the precipitated DCU was filtered off, washed with CH₂Cl₂, and the residue was subjected to ¹H NMR analysis as a solution in chloroform-*d* (ester of **3**) or benzene-*d*₆ (ester of **4**). Integrals of signals at 4.695 and 4.684 ppm (*H*-C-OMe) and 3.294 and 3.254 ppm (CH₃O-C-H) in the spectra of *O*-methylmandelate of **3** and 4.618 and 4.586 ppm (*H*-C-OMe) and 3.176 and 3.144 ppm (CH₃O-C-H) in the spectra of *O*-methylmandelate of **4** were selected for calculation of e.e.

4.2.2. Method 2: by HPLC. Retention times: *t*_R(*R*)-**3**=15.16 min, *t*_R(*S*)-**3**=11.40 min; *t*_R(*R*)-**4**=6.34 min, *t*_R(*S*)-**4**=7.55 min.

4.3. Phosphonylation of aldehydes (*S*)-**1** and (*S*)-**2**

4.3.1. Method 1: with dimethyl(trimethylsilyl)phosphite. To a solution of a crude aldehyde (*S*)-**2** [prepared from (*S*)-**4** (1.66 g, 7.00 mmol)] in CH₂Cl₂ (3 mL), (MeO)₂POTMS (1.34 mL, 7.00 mmol) was added at –20°C. The reaction mixture was left at –20°C for 24 h. The solvent was evaporated, and the residue was dissolved in 1 M Bu₄NF in THF (8.4 mL, 8.4 mmol). After 1 h at rt the solvent was evaporated, the crude product was dissolved in CH₂Cl₂ (10 mL), washed with water (2×5 mL) and dried over MgSO₄. Column chromatography on silica gel gave a 83:17 mixture of (1*S*,2*S*)-**9a** and (1*R*,2*S*)-**9b** (1.92 g, 80%).

In the same way, from crude aldehyde (*S*)-**1** [prepared from (*S*)-**3** (0.241 g, 1.00 mmol)] and (MeO)₂POTMS (0.19 mL, 1.00 mmol), a 87:13 mixture of (1*S*,2*S*)-**10a** and (1*R*,2*S*)-**10b** (0.216 g, 62%) was obtained after chromatography on a silica gel column.

4.3.2. Method 2: with lithium *O,O*-dimethyl phosphite. Crude aldehyde (*S*)-**2** [prepared from (*S*)-**4** (1.42 g, 6.00 mmol)] was subjected to the reaction with lithium *O,O*-dimethyl phosphite (6.00 mmol) at –70°C as described in Ref. 12. After chromatography on a silica gel column a 75:25 mixture of the phosphonates (1*S*,2*S*)-**9a** and (1*R*,2*S*)-**9b** (1.347 g, 80%) was obtained.

In the same way, from crude aldehyde (*S*)-**1** [prepared from (*S*)-**3** (0.668 g, 2.77 mmol)] and lithium *O,O*-dimethyl phosphite (2.77 mmol), a 1:1 mixture of the phosphonates (*1S,2S*)-**10a** and (*1R,2S*)-**10b** (0.490 g, 51%) was obtained after chromatography on a silica gel column.

4.4. Phosphonates (*1S,2S*)-**9a** and (*1S,2S*)-**10a**

4.4.1. Benzoate, 11a. A 3:1 mixture of (*1S,2S*)-**9a** and (*1R,2S*)-**9b** (1.025 g, 2.968 mmol) was esterified with benzoyl chloride as described earlier¹⁴ to give crude product (0.666 g, 83%), which was purified on a silica gel column followed by crystallisation. The benzoate (*1S,2S*)-**11a** was obtained in 30% yield (0.392 g). Mp 93.0–93.8°C.

4.4.2. Phosphonate (*1S,2S*)-10a.** The benzoate (*1S,2S*)-**11a** (0.130 g, 0.289 mmol) was treated with 3.7 M HCl–AcOEt as described earlier¹⁴ to afford (*1S,2S*)-**10a** (0.082 g, 82%). Mp. 117–118°C. Retention time: $t_R = 21.92$ min.**

Retention times for the racemic mixture: $t_R(1R,2R)$ -**10a** = 16.73 min, $t_R(1S,2S)$ -**10a** = 21.95 min.

4.4.3. Phosphonate (*1S,2S*)-9a.** The benzoate (*1S,2S*)-**11a** (0.075 g, 0.167 mmol) was dissolved in methanol (0.5 mL) and treated with aqueous ammonia (25%, 1 mL) for 24 h.¹⁴ After column chromatography on a silica gel column, (*1S,2S*)-**9a** (0.022 g, 40%) was obtained as a colourless oil. Retention time: $t_R = 8.30$ min.**

Retention times for the racemic mixture: $t_R(1R,2R)$ -**9a** = 10.04 min, $t_R(1S,2S)$ -**9a** = 8.86 min.

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References

- Jurczak, J.; Gołębiowski, A. *Chem. Rev.* **1989**, *89*, 149–164.
- Veeresa, G.; Datta, A. *Tetrahedron Lett.* **1998**, *39*, 3069–3070.
- Veeresa, G.; Datta, A. *Tetrahedron Lett.* **1998**, *39*, 119–3070.
- Matsunga, N.; Harada, H.; Aoyama, T.; Shioiri, T. *Heterocycles* **1992**, *33*, 235–255.
- Lee, K.-Y.; Kim, Y.-H.; Park, M.-S.; Oh, C.-Y.; Ham, W.-H. *J. Org. Chem.* **1999**, *64*, 9450–9458.
- Schade, W.; Reissig, H.-U. *J. Prakt. Chem.* **1999**, *341*, 685–686.
- Ambroise, L.; Jackson, R. F. W. *Tetrahedron Lett.* **1996**, *37*, 2311–2314.
- Barco, A.; Benetti, S.; Risi, C. D.; Pollini, G. P.; Romagnoli, R.; Zanirato, V. *Tetrahedron Lett.* **1994**, *35*, 9289–9292.
- Dondoni, A.; Perrone, D.; Semola, T. *Synthesis* **1995**, 181–186.
- Denis, J.-N.; Correa, A.; Greene, A. E. *J. Org. Chem.* **1991**, *56*, 6939–6942.
- Jayasinghe, L. R.; Datta, A.; Ali, S. M.; Zygmunt, J.; Vander Velde, D. G.; Georg, G. I. *J. Med. Chem.* **1994**, *37*, 2981–2984.
- Wróblewski, A. E.; Piotrowska, D. G. *Tetrahedron* **1998**, *54*, 8123–8132.
- Wróblewski, A. E.; Piotrowska, D. G. *Tetrahedron: Asymmetry* **1999**, *10*, 2037–2044.
- Wróblewski, A. E.; Piotrowska, D. G. *Tetrahedron: Asymmetry* **2000**, *11*, 2615–2624.
- Kawano, T.; Ogawa, T.; Islam, S. Md.; Ueda, I. *Heterocycles* **2000**, *52*, 1279–1295.
- Alexakis, A.; Lensen, N.; Mangeney, P. *Synlett* **1991**, 625–626.
- Morita, T.; Nagasawa, Y.; Yahiro, S.; Matsunaga, H.; Kunieda, T. *Org. Lett.* **2001**, *3*, 897–899.
- Myers, A. G.; Kung, D. W.; Zhong, B. *J. Am. Chem. Soc.* **2000**, *122*, 3236–3237.
- Myers, A. G.; Zhong, B.; Kung, D. W.; Movassaghi, M.; Lanman, B. A.; Kwon, S. *Org. Lett.* **2000**, *2*, 3337–3340.
- Myers, A. G.; Zhong, B.; Movassaghi, M.; Kung, D. W.; Lanman, B. A.; Kwon, S. *Tetrahedron Lett.* **2000**, *41*, 1359–1362.
- Chang, Z. Y.; Coats, R. M. *J. Org. Chem.* **1990**, *55*, 3464–3474.
- Lewandowicz, A.; Lipiński, J.; Siedlecka, R.; Skarzewski, J.; Baert, F. *Tetrahedron* **1998**, *54*, 6571–6586.
- Mancuso, A. J.; Huang, S.-L.; Swern, D. *J. Org. Chem.* **1978**, *43*, 2480–2482.
- Mancuso, A. J.; Swern, D. *Synthesis* **1981**, 165–185.
- Dess, D. B.; Martin, J. C. *J. Org. Chem.* **1983**, *48*, 4155–4156.
- Dess, D. B.; Martin, J. C. *J. Am. Chem. Soc.* **1991**, *113*, 7277–7287.
- Ireland, R. E.; Liu, L. *J. Org. Chem.* **1993**, *58*, 2899.
- Meyer, S. D.; Schreiber, S. L. *J. Org. Chem.* **1994**, *59*, 7549–7552.
- Piotrowska, D. G.; Hałajewska-Wosik, A.; Wróblewski, A. E. *Synth. Commun.* **2000**, *30*, 3935–3940.